Journal of Life Science and Applied Research (JLSAR) E-ISSN: 2959-8036



CADMIUM BIOREMEDIATION BY BREVIBACILLUS SPP: A MINIREVIEW

N. M. T. Jebril ២

Article Info:

Received: Jul. 11, 2024 Revised: Aug. 09, 2024 Accepted: Sep. 18, 2024 Published: Dec. 31, 2024

DOI:

https://doi.org/10.59807/jlsar.v 5i2.98

How to Cite:

N. M. T. . Jebril, "CADMIUM BIOREMEDIATION BY BREVIBACILLUS SPP: A MINIREVIEW", JLSAR, vol. 5, no. 2, pp. 60–69, Dec. 2024.

Available From: https://www.jlsar.com/index.php/journal/article/view/98



Copyright: © 2024 by the authors. Submitted for possible open-access publication under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecom-mons.org/licenses/by/4.0/).



 Department of Biology, College of Sciences for Women, University of Babylon, Iraq.
 * Corresponding author: N. M. T. Jebril, Department of Biology, College of Sciences for Women, University of Babylon, Iraq. Email: <u>nadia.tawfiq@uobabylon.edu.iq</u>

Abstract: This review examines the potential of Brevibacillus species in cadmium bioremediation, a critical environmental challenge due to the harmful effects of cadmium contamination. The review highlights the limitations of traditional chemical methods, such as precipitation, which are inefficient at low cadmium concentrations and costly. Instead, biological methods are increasingly favored for their cost-effectiveness, sustainability, and efficiency, particularly at dilute concentrations. The review focuses on the genus Brevibacillus, a group of bacteria known for their diverse environmental habitats and biotechnological applications. Specific species of Brevibacillus have demonstrated the ability to degrade pollutants and accumulate heavy metals, including cadmium. For example, B. agri has been identified as having significant potential for cadmium tolerance and bioaccumulation. The mechanisms employed by these bacteria include biosorption, bioaccumulation, and extracellular uptake, making them effective candidates for bioremediation strategies. The review emphasizes that while several Brevibacillus species have been applied in cadmium removal, further studies are necessary to fully explore the potential of B. agri and other species in various contaminated environments. The findings underscore the importance of leveraging microbial processes for environmental cleanup, especially in areas affected by heavy metal pollution.

Keywords: Adsorption; Bioaccumulation; Bioremediation; Brevibacillus agri; Cadmium.

1. Introduction

A variety of combine physiochemical or chemical techniques is available for immobilization of cadmium contaminant from water, including those that use chemical oxidation, chemical precipitation, ion exchange, and surfactant-enhanced adsorption. Although the most removal cadmium (II) technique is using precipitation there are certain disadvantages of the chemical precipitation techniques. The chemical precipitation technology is not cost-effective for the re-

moval of cadmium (II) at a low concentration from contaminated environments [1]. Surfactant-enhanced adsorption technology has applied widely to cadmium adsorption using a variety of surfactants such as agricultural wastes, granular activated carbon, industrial wastewater [2]. Biological methods; however, have been sought as an alternative strategy. Hence, there is a demand for safe, and effective ways to remove cadmium (II) contaminant from the environment, considering alternative treatment of cadmium contaminated groundwater is extremely important for public health. Biological treatment techniques are considered one of the remediation techniques that increasingly gaining attention as an alternative technology because of their potential uses for providing simplicity, efficiency, and cost-effective technology for cadmium remediation especially at a dilute concentration [3]. These technologies exploit biological process by certain plant, alga, fungi, yeast, and *Bacteria* to bioremediate cadmium. The biological mechanisms for the removal of cadmium from water by living microorganism are via biosorption, extracellular or intercellular uptake, accumulation, complexation, oxidation, and precipitation [4]. Reviews of [5], [6], [7], [8] included the capacities of some of bacteria and plant, alga, fungi, and yeast species that used previously for cadmium removal. Bioremediation techniques are more sustainable, and economical technologies even some bioremediation techniques in compared to physical, and physiochemical techniques [9]. The first practice of bacteria for Cd bioremediation was explored in species of *Citrobacter* [10], [11], [12]. Subsequent these expectations, this *Citrobacter* was too considered for this purpose in different conditions [11], [13]. And later, more amendment method was applied for using *Citrobacter* sp. in Cd bioremediation such as in solutions supplemented with phosphatase substrate [14], [15]. Also, *Citrobacter* MCM B-181 was used for Cd bioremediation [16].

2. Cadmium (II) bioremediation

Bacteria have bioremediation mechanisms by which cadmium (II) can be taken up, and remove cadmium (II) from the solution. An understanding of these mechanisms is important to utilise the selectivity, and efficiency of the optimisation of the bioremediation process [17], [18], [19]. A reviews of [5], [6] described the catabolic landscape and physiochemical mechanisms.

3. Review bacterium Brevibacillus

The *Brevibacillus* spp. are forming a group thermophilic, psychrophilic, acidophilic, alkalophilic or halophilic, Gram positive, negative or variable, and rod-shaped *Bacteria*. The members of the genus are either strictly aerobic or facultative anaerobic, and optimally grow within the range of temperature 30 °C to 45 °C [20]. Changes in bacterial taxonomy have been reflected in the classification scheme used to define on the basis of 16S rDNA gene sequence analysis which was, up until 1996. The genus *Brevibacillus* was first described by [21], and reclassified as a species belonging to the novel genus *Brevibacillus* by [22]. The shift toward a phylogenetic classification system dram antically affected the biological identity of the genus *Brevibacillus*, which was a relatively large and important group of Gam-positive. Currently, the genus *Brevibacillus* includes 20 species allotted to the novel genus *Brevibacillus*; namely: *B. brevis*, *B. agri*, *B. borstelensis*, *B. centrosporus*, *B. choshinensis*, *B. formosus*, *B. ginsengisoli*, *B. invocatus*, *B. laterosporus*, *B. levickii*, *B. limnophilus*, *B. parabrevis*, *B. reuszeri*, *B. thermoruber*, *B. aydinogluensis*, *B. fluminis*, *B. fulvus*, *B. ginsengisoli*, *B. massiliensis*, *B. nitrificans*, and *B. panacihumi* [23], [24], [25], [26], [27], [28], [29], [30]. The general features of species from genus *Brevibacillus* has been reviewed by [31].

4. Brevibacillus spp. in polluted environments

The species of *Brevibacillus* have been isolated from the diverse environmental habitats, and geographical locations; this includes waste compost, oil fields, hot water springs, food, and food processing environments, volcanoes, and hydrothermal marine vents, pharmaceutical products, heat treatment of specimens, human clinical specimens, and from human illness or with insect pathogenicity.

5. Biotechnological applications of Brevibacillus spp. used in bioremediation

Regarding to the biotechnological applications, different species of *Brevibacillus* were used previously for different bioremediation studies. Table 1 summaries the literary of some *Brevibacillus* species isolated in previous studies with their ecological distribution, and biotechnological applications.

Some *Brevibacillus* spp. have been applied in biodegradation of pollutants such as biodegradation of lowdensity of polyethene by *B. parabrevis* [32], and of triphenyltin by *B. brevis*. Additionally, species of *Brevibacillus* sp. have been found to be able to biodegrade of keratins [33] or oil recovery as has been studied in *B. brevis* [34]. *Brevibacillus* spp. also have the ability to act as a candidate biocontrol agents due to their capacities to produce extracellular neutral protease. The biocontrol potential of this species has been reported against nematode control [35], insects, *Lepidoptera* and *Coleoptera*, *Musca domestica*, and *Aedes aegypti* [36]. In addition to its pathogenicity against invertebrates, different strains of *B. laterosporus* showed a broad-spectrum antimicrobial activity especially against Bacteria, fungi, and mycobacteria [37], [38]. Therefore, the wide ranges of using the *Brevibacillus* species were supported the potential used of this bacterium for cadmium removal.

<i>Brevibacillus</i> species	Environmental source	Applications	Reference
B. levickii	Geothermal soils of northern Victoria L,	Possessed an uptake system specific for	[37]
	and, Antarctica	L-glutamic acid	
B. invocatus	An outbreak of water-borne illness in four	Associated with an outbreak of	[30]
	Swedish towns	waterborne illness	
Brevibacillus sp.	Coomassie brilliant blue- polluted soil	Degrade toluidine blue dye (TB)	[39]
B. brevis	Nickel-contaminated soil	Nickel tolerant	[40]
B. brevis	Cadmium-contaminated soil,	Cadmium tolerance	[41]
	Nagyhörcsök, Hungary		
B. brevis	Cadmium-polluted soil	Cadmium tolerance	[41]
B. brevis	Cadmium-contaminated soils	Cadmium bioaccumulation ability	[42]
B. laterosporus	Soil sediments, near a wastewater	Dye-decolorization	[43]
	effluent outlet of a local cotton textile		
	factory, Nakhonpathom, Thailand		
B. brevis	Soil sediment, Guiyu, Guangdong	Biosorption , and biodegradation of	[44]
	Province, China	triphenyltin	
[45]	Lead and cadmium-polluted soil	Lead and cadmium absorbed	[45]
B. laterosporus	Microbial type culture collection,	Biodegradation of dissimilar dyes	[46]
	Chandigarh, India		
B. laterosporus	Microbial type culture collection,	Biodegradation , and detoxification of	[47]
	Chandigarh, India	textile dye	
B. brevis	Soil, saontalpara, Haringhata, Nadia,	Biotransformation, and bioaccumulation	[48]
	India	of arsenic	
Brevibacillus sp.	Arsenic-contaminated soil	Arsenic bioremediation	[49]
B. brevis	Wastewater Abeokuta, Ogun state, Nigeria	Removal of chromate	[50]
Brevibacillus sp.	Indonesia's hot Springs	Alkaline protease	[51]
B. agri	Engineering B.agri dih-ydropyrimidinase	Production of l-homophenylalanine	[52]
Brevibacillus sp.	Diyadin hot Spring, Agri, Turkey	Removal of textile dyes	[53]
Brevibacillus sp.	Rhizosphere, arsenic- contaminated soil,	Bioremediation of arsenic	[54]
	Nadia, India		
B. borstelensis	Hasanabdal hot spring water, Ercis, Van,	Fruit juice , and oil extraction	[55]
	Turkey		
B. thermoruber	Hot springs naturally enriched with	Reduced ionic mercury	[56]
	mercury, Mount Amiata, Tuscany, Italy,		

Table 1: Summaries of ecological distribution, and biotechnological applications of some *Brevibacillus* species, previously isolated.

B. brevis	Marine sediments, Guiyu, Guangdong,	Biosorption and biodegradation of	[57]
	China	pyrene	
B. thermoruber	Feather-contaminated soil	Degradation of native feathers	[58]
B. brevis	Soil, Scheyern, Bavaria, Germany	Degradation of chiral fungicides metalaxyl and furalaxyl	[58]
B. nitrificans	Microbiological agent	Nitrogen removal in sewage treatment tanks	[59]
B. laterosporus		Degradation of polyvinyl alcohol	[60]
B. borstelensis GIGAN1	A river sludge in Guangzhou, China	Biodeodorization of thioanisole	[61]
B. parabrevis	Water bodies located near to the carpet industries, Bhadohi, India	Bioremediation of Congo red dye	[62]
B. laterosporus	Marine sediments, Paradip port, Odisha,	Reduction of chromium	[63]
CrRPSD40	India		
B. agri DH-1	Soil	Remove dichlorobenzene	[64]
B. invocatus C19	Egyptian Coke	Desulfurization of dibenzothiophene	[65]
B.agri CAT37	Gas-washing wastewaters	Biodegradation of malodorous thiols	[66]
B. laterosporus MTCC 2298		Biodegradation of malachite green, triphenylmethane dyes	[67]
B. panacihumi ZB1		Biological treatment, COD	[68]
B. brevis		Bioremediation of triphenyl phosphate	[69]
B. parabrevis MTCC 12105	Pulp , and paper mill sludge	Pulp , and paper effluent degradation	[70]
B. agri C15	Cadmium-contaminated soil	Cadmium tolerance, cadmium bioaccumulation ability	[71], [72], [73]
B. agri C15 Cd ^R	Ultraviolet light mutagenesis of <i>B. agri</i> C15	Cadmium tolerance, cadmium bioaccumulation ability	[73], [74], [75]

6. Conclusion

This In conclusion, Brevibacillus spp. expresses its attitudes like nickel, lead, chro-mate, arsenic and cadmium tolerant, biodegradation of pollutants and biocontrol agents. Brevibacillus spp. were isolated previously from cadmium-contaminated environments, and used for enhancing the uptake of cadmium by plant of contaminated soils. But none of any study has been used B. agri for cadmium removal, especially. To our knowledge, there is no study which identified the cadmium resistance of the B. agri. The above-mentioned features were confirming and concluded that B. agri could be one of the most appropriate bacterium or more right choice for cadmium bioremediation.

Supplementary Materials:

No Supplementary Materials.

Author Contributions:

N. M. T. Jebril ; methodology, writing—original draft preparation, N. M. T. Jebril ; writing—review and editing, N. M. T. Jebril ; paraphrasing. The author has read and agreed to the published version of the manuscript.

Funding:

This research received no external funding.

Institutional Review Board Statement:

The study was conducted in accordance with the protocol authorized by , College of Sciences for Women, University of Babylon, Iraq.

Informed Consent Statement:

No Informed Consent Statement.

Data Availability Statement:

No Data Availability Statement.

Conflicts of Interest:

The authors declare no conflict of interest.

Acknowledgments:

Many thanks for Dr Rich Boden, University of Plymouth, England for his information and instructions.

Disclaimer/Journal's Note:

The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of JLSAR and/or the editor(s). JLSAR and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

7. References

- R. A. Wuana and F. E. Okieimen, "Heavy Metals in Contaminated Soils: A Review of Sources, Chemistry, Risks and Best Available Strategies for Remediation," *ISRN Ecol*, vol. 2011, 2011, doi: 10.5402/2011/402647.
- [2] M. Kazemipour, M. Ansari, S. Tajrobehkar, M. Majdzadeh, and H. R. Kermani, "Removal of lead, cadmium, zinc, and copper from industrial wastewater by carbon developed from walnut, hazelnut, almond, pistachio shell, and apricot stone," *J Hazard Mater*, vol. 150, no. 2, 2008, doi: 10.1016/j.jhazmat.2007.04.118.
- [3] J. Tausz and P. Donath, "Über die Oxydation des Wasserstoffs und der Kohlenwasserstoffe mittels Bakterien," *Hoppe Seylers Z Physiol Chem*, vol. 190, no. 3–6, 1930, doi: 10.1515/bchm2.1930.190.3-6.141.
- [4] B. M. In, *Emerging technology for bioremediation of metals (Vol. 2). CRC Press.* 1994.
- [5] N. Mahmoud and T. Jebril, "Systems biology design of the microbe toward the bioremediation of cadmium: a review," Article in International Journal of Psychosocial Rehabilitation, 2020, doi: 10.37200/IJPR/V24I8/PR280906.
- [6] N. Jebril, R. Boden, and C. Braungardt, "Cadmium resistant bacteria mediated cadmium removal: a systematic review on resistance, mechanism and bioremediation approaches," in *IOP Conference Series: Earth and Environmental Science*, 2022. doi: 10.1088/1755-1315/1002/1/012006.
- [7] N. Jebril, R. Boden, and C. Braungardt, "THE GROUNDWATER CONTAMINATED WITH CAD-MIUM: A SYSTEMATIC REVIEW," ANBAR JOURNAL OF AGRICULTURAL SCIENCES, vol. 20, no. 2, 2022, doi: 10.32649/ajas.2022.176912.
- [8] N. M. T. Jebril, "Nanobiopolymer: Potential Applications in Bioremediation of Cadmium Contaminated Water," *Iraqi Journal of Industrial Research*, vol. 10, no. 2, 2023, doi: 10.53523/ijoirvol10i2id290.

[9]	M. Tyagi, M. M. R. da Fonseca, and C. C. C. R. de Carvalho, "Bioaugmentation and biostimulation
	strategies to improve the effectiveness of bioremediation processes," 2011. doi: 10.1007/s10532-010-9394-4.
[10]	L. E. Macaskie and A. C. R. Dean, "Cadmium accumulation by a Citrobacter sp.," <i>J Gen Microbiol</i> , vol. 130, no. 1, 1984, doi: 10.1099/00221287-130-1-53.
[11]	L. J. Michel, L. E. Macaskie, and A. C. R. Dean, "Cadmium accumulation by immobilized cells of
	a Citrobacter sp. using various phosphate donors," <i>Biotechnol Bioeng</i> , vol. 28, no. 9, 1986, doi:
	10.1002/bit.260280910.
[12]	L. E. Macaskie and A. C. R. Dean, "Heavy metal accumulation by immobilized cells of a Citrobac-
	ter sp.," <i>Biotechnol Lett</i> , vol. 6, no. 2, 1984, doi: 10.1007/BF00127292.
[13]	L. E. Macaskie, J. M. Wates, and A. C. R. Dean, "Cadmium accumulation by a Citrobacter sp.
	immobilized on gel and solid supports: Applicability to the treatment of liquid wastes containing
	heavy metal cations," <i>Biotechnol Bioeng</i> , vol. 30, no. 1, 1987, doi: 10.1002/bit.260300110.
[14]	L. E. Macaskie, A. C. R. Dean, and A. K. Cheetham, "Cadmium accumulation by a Citrobacter sp.:
	The chemical nature of the accumulated metal precipitate and its location on the bacterial cells,"
	J Gen Microbiol, vol. 133, no. 3, 1987, doi: 10.1099/00221287-133-3-539.
[15]	L. E. Macaskie, K. M. Bonthrone, and D. A. Rouch, "Phosphatase-mediated heavy metal accumu-
	lation by a Citrobacter sp. and related enterobacteria," FEMS Microbiol Lett, vol. 121, no. 2, 1994,
	doi: 10.1111/j.1574-6968.1994.tb07090.x.
[16]	P. R. Puranik and K. M. Paknikar, "Biosorption of lead, cadmium, and zinc by Citrobacter strain
	MCM B-181: Characterization studies," <i>Biotechnol Prog</i> , vol. 15, no. 2, 1999, doi: 10.1021/bp990002r.
[17]	T. J. Beveridge and R. G. E. Murray, "Sites of metal deposition in the cell wall of Bacillus subtilis,"
	J Bacteriol, vol. 141, no. 2, 1980, doi: 10.1128/jb.141.2.876-887.1980.
[18]	M. Tsezos and B. Volesky, "The mechanism of uranium biosorption by Rhizopus arrhizus," Bio-
	technol Bioeng, vol. 24, no. 2, 1982, doi: 10.1002/bit.260240211.
[19]	B. Volesky, J. Weber, and J. M. Park, "Continuous-flow metal biosorption in a regenerable Sar-
	gassum column," Water Res, vol. 37, no. 2, 2003, doi: 10.1016/S0043-1354(02)00282-8.
[20]	D. H. Bergey and D. R. Boone, Bergey's Manual Of Systematic Bacteriology, Volume 2, Part 3, vol. 2.
	2001.
[21]	E. KLEIN, "System der Bakterien," Nature, vol. 61, no. 1585, 1900, doi: 10.1038/061464a0.
[22]	O. Shida, H. Takagi, K. Kadowaki, and K. Komagata, "Proposal for two new genera, Brevibacillus
	gen. nov. and Aneurinibacillus gen. nov.," Int J Syst Bacteriol, vol. 46, no. 4, 1996, doi:
	10.1099/00207713-46-4-939.
[23]	P. Hugon et al., "Non-contiguous finished genome sequence and description of brevibacillus mas-
	siliensis sp. nov," Stand Genomic Sci, vol. 8, no. 1, 2013, doi: 10.4056/sigs.3466975.
[24]	V. Sharma, P. K. Singh, S. Midha, M. Ranjan, S. Korpole, and P. B. Patil, "Genome sequence of
	Brevibacillus laterosporus strain GI-9," 2012. doi: 10.1128/JB.06659-11.
[25]	M. J. Choi, J. Y. Bae, K. Y. Kim, H. Kang, and C. J. Cha, "Brevibacillus fluminis sp. nov., isolated
	from sediment of estuarine wetland," 2010. doi: 10.1099/ijs.0.012351-0.
[26]	M. K. Kim, S. Sathiyaraj, R. K. Pulla, and D. C. Yang, "Brevibacillus panacihumi sp. nov., a β -
	glucosidase-producing bacterium," Int J Syst Evol Microbiol, vol. 59, no. 5, 2009, doi:
	10.1099/ijs.0.001248-0.
[27]	N. Parvez, L. K. Cornelius, and R. Fader, "Brevibacillus brevis peritonitis," American Journal of the
	Medical Sciences, vol. 337, no. 4, 2009, doi: 10.1097/MAJ.0b013e3181891626.

- [28] S. H. Baek, W. T. Im, H. W. Oh, J. S. Lee, H. M. Oh, and S. T. Lee, "Brevibacillus ginsengisoli sp. nov., a denitrifying bacterium isolated from soil of a ginseng field," *Int J Syst Evol Microbiol*, vol. 56, no. 11, 2006, doi: 10.1099/ijs.0.64382-0.
- [29] K. Goto, R. Fujita, Y. Kato, M. Asahara, and A. Yokota, "Reclassification of Brevibacillus brevis strains NCIMB 13288 and DSM 6472 (=NRRL NRS-887) as Aneurinibacillus danicus sp. nov. and Brevibacillus limnophilus sp. nov," *Int J Syst Evol Microbiol*, vol. 54, no. 2, 2004, doi: 10.1099/ijs.0.02906-0.
- [30] N. A. Logan *et al.*, "Polyphasic identification of Bacillus and Brevibacillus strains from clinical, dairy and industrial specimens and proposal of Brevibacillus invocatus sp. nov," *Int J Syst Evol Microbiol*, vol. 52, no. 3, 2002, doi: 10.1099/ijs.0.02081-0.
- [31] A. K. Panda, S. S. Bisht, S. DeMondal, N. Senthil Kumar, G. Gurusubramanian, and A. K. Panigrahi, "Brevibacillus as a biological tool: A short review," 2014. doi: 10.1007/s10482-013-0099-7.
- [32] R. Pramila, "Brevibacillus parabrevis, Acinetobacter baumannii and Pseudomonas citronellolis -Potential candidates for biodegradation of low density polyethylene (LDPE)," *Journal of Bacteriol*ogy Research, vol. 4, no. 1, 2012, doi: 10.5897/jbr12.003.
- [33] N. Z. Jaouadi *et al.*, "Biochemical and Molecular Characterization of a Serine Keratinase from Brevibacillus brevis US575 with Promising Keratin-Biodegradation and Hide-Dehairing Activities," *PLoS One*, vol. 8, no. 10, 2013, doi: 10.1371/journal.pone.0076722.
- [34] S. Ebrahimi and A. A. Sepahi, "Exopolysacarid production by strain of Brevibacillus brevis: Potential applications in the treatment of hydrocarbons pollution and use in microbial enhance oil recovery (MEOR)," 2008.
- [35] B. Tian, N. Li, L. Lian, J. Liu, J. Yang, and K. Q. Zhang, "Cloning, expression and deletion of the cuticle-degrading protease BLG4 from nematophagous bacterium Brevibacillus laterosporus G4," *Arch Microbiol*, vol. 186, no. 4, 2006, doi: 10.1007/s00203-006-0145-1.
- [36] L. Ruiu, I. Floris, A. Satta, and D. J. Ellar, "Toxicity of a Brevibacillus laterosporus strain lacking parasporal crystals against Musca domestica and Aedes aegypti," *Biological Control*, vol. 43, no. 1, 2007, doi: 10.1016/j.biocontrol.2007.07.002.
- [37] D. D. C. Carvalho, M. Lobo Junior, I. Martins, P. W. Inglis, and S. C. M. Mello, "Biological control of fusarium oxysporum f. sp. phaseoli by trichoderma harzianum and its use for common bean seed treatment," *Trop Plant Pathol*, vol. 39, no. 5, 2014, doi: 10.1590/S1982-56762014000500005.
- [38] T. Arumugam, P. Senthil Kumar, R. V. Hemavathy, V. Swetha, and R. Karishma Sri, "Isolation, structure elucidation and anticancer activity from Brevibacillus brevis EGS 9 that combats Multi Drug Resistant actinobacteria," *Microb Pathog*, vol. 115, 2018, doi: 10.1016/j.micpath.2017.12.061.
- [39] H. A. Alhassani, M. A. Rauf, and S. S. Ashraf, "Efficient microbial degradation of Toluidine Blue dye by Brevibacillus sp.," *Dyes and Pigments*, vol. 75, no. 2, 2007, doi: 10.1016/j.dyepig.2006.06.019.
- [40] A. Vivas, B. Biró, T. Németh, J. M. Barea, and R. Azcón, "Nickel-tolerant Brevibacillus brevis and arbuscular mycorrhizal fungus can reduce metal acquisition and nickel toxicity effects in plant growing in nickel supplemented soil," *Soil Biol Biochem*, vol. 38, no. 9, 2006, doi: 10.1016/j.soilbio.2006.04.020.
- [41] A. Vivas, I. Vörös, B. Biró, E. Campos, J. M. Barea, and R. Azcón, "Symbiotic efficiency of autochthonous arbuscular mycorrhizal fungus (G. mosseae) and Brevibacillus sp. isolated from cadmium polluted soil under increasing cadmium levels," *Environmental Pollution*, vol. 126, no. 2, 2003, doi: 10.1016/S0269-7491(03)00195-7.

- [42] A. Vivas, J. M. Barea, and R. Azcón, "Brevibacillus brevis isolated from cadmium- or zinc-contaminated soils improves in vitro spore germination and growth of Glomus mosseae under high Cd or Zn concentrations," *Microb Ecol*, vol. 49, no. 3, 2005, doi: 10.1007/s00248-004-0044-4.
- [43] W. Lang *et al.*, "Characterization of a new oxygen-insensitive azoreductase from Brevibacillus laterosporus TISTR1911: Toward dye decolorization using a packed-bed metal affinity reactor," *Bioresour Technol*, vol. 150, 2013, doi: 10.1016/j.biortech.2013.09.124.
- [44] J. Ye, H. Yin, H. Peng, J. Bai, D. Xie, and L. Wang, "Biosorption and biodegradation of triphenyltin by Brevibacillus brevis," *Bioresour Technol*, vol. 129, 2013, doi: 10.1016/j.biortech.2012.11.076.
- [45] J. M. Ruiz-Lozano and R. Azcón, "Brevibacillus, Arbuscular Mycorrhizae and Remediation of Metal Toxicity in Agricultural Soils," 2011. doi: 10.1007/978-3-642-19577-8_12.
- [46] M. B. Kurade, T. R. Waghmode, and S. P. Govindwar, "Preferential biodegradation of structurally dissimilar dyes from a mixture by Brevibacillus laterosporus," J Hazard Mater, vol. 192, no. 3, 2011, doi: 10.1016/j.jhazmat.2011.07.004.
- [47] M. B. Kurade, T. R. Waghmode, R. V. Khandare, B. H. Jeon, and S. P. Govindwar, "Biodegradation and detoxification of textile dye Disperse Red 54 by Brevibacillus laterosporus and determination of its metabolic fate," J Biosci Bioeng, vol. 121, no. 4, 2016, doi: 10.1016/j.jbiosc.2015.08.014.
- [48] S. Banerjee, "Biotransformation and bioaccumulation of arsenic by Brevibacillus brevis isolated from arsenic contaminated region of West Bengal," *IOSR J Environ Sci Toxicol Food Technol*, vol. 3, no. 1, 2013, doi: 10.9790/2402-0310110.
- [49] I. Mallick, S. T. Hossain, S. Sinha, and S. K. Mukherjee, "Brevibacillus sp. KUMAs2, a bacterial isolate for possible bioremediation of arsenic in rhizosphere," *Ecotoxicol Environ Saf*, vol. 107, 2014, doi: 10.1016/j.ecoenv.2014.06.007.
- [50] P. Wani, A. Olamide, N. Rafi, S. Wahid, I. Wasiu, and O. Sunday, "Sodium Alginate/Polyvinyl Alcohol Immobilization of Brevibacillus brevis OZF6 Isolated from Waste Water and Its Role in the Removal of Toxic Chromate," *Br Biotechnol J*, vol. 15, no. 1, 2016, doi: 10.9734/bbj/2016/27341.
- [51] S. Wang, X. Lin, X. Huang, L. Zheng, and D. S. Zilda, "Screening and characterization of the alkaline protease isolated from PLI-1, a strain of Brevibacillus sp. collected from Indonesia's hot springs," *Journal of Ocean University of China*, vol. 11, no. 2, 2012, doi: 10.1007/s11802-012-1845-6.
- [52] C. K. Lo *et al.*, "Engineering of the critical residues at the stereochemistry-gate loops of Brevibacillus agri dihydropyrimidinase for the production of l-homophenylalanine," *Process Biochemistry*, vol. 44, no. 3, 2009, doi: 10.1016/j.procbio.2008.11.005.
- [53] C. Bozoglu, A. Adiguzel, H. Nadaroglu, D. Yanmis, and M. Gulluce, "Pur ification and Char acter ization of Laccase fr om newly isolated Ther m ophilic Br eviba cillus sp. (Z1) and its applications in r em oval of Textile D yes," 2013.
- [54] I. Mallick and S. K. Mukherjee, "Bioremediation potential of an arsenic immobilizing strain Brevibacillus sp. KUMAs1 in the rhizosphere of chilli plant," *Environ Earth Sci*, vol. 74, no. 9, 2015, doi: 10.1007/s12665-015-4686-y.
- [55] N. Demir *et al.*, "Purification and characterization of an alkaline pectin lyase produced by a newly isolated brevibacillus borstelensis (p35) and its applications in fruit juice and oil extraction," *European Food Research and Technology*, vol. 239, no. 1, 2014, doi: 10.1007/s00217-014-2198-8.
- [56] A. D. Chatziefthimiou, M. Crespo-Medina, Y. Wang, C. Vetriani, and T. Barkay, "The isolation and initial characterization of mercury resistant chemolithotrophic thermophilic bacteria from mercury rich geothermal springs," *Extremophiles*, vol. 11, no. 3, 2007, doi: 10.1007/s00792-007-0065-2.

- [57] L. Liao *et al.,* "Biosorption and biodegradation of pyrene by Brevibacillus brevis and cellular responses to pyrene treatment," *Ecotoxicol Environ Saf,* vol. 115, 2015, doi: 10.1016/j.ecoenv.2015.02.015.
- [58] L. Sulimma, A. Bullach, S. Kusari, M. Lamshöft, S. Zühlke, and M. Spiteller, "Enantioselective degradation of the chiral fungicides metalaxyl and furalaxyl by Brevibacillus brevis," *Chirality*, vol. 25, no. 6, 2013, doi: 10.1002/chir.22158.
- [59] F. Takebe, K. Hirota, Y. Nodasaka, and I. Yumoto, "Brevibacillus nitrificans sp. nov., a nitrifying bacterium isolated from a microbiological agent for enhancing microbial digestion in sewage treatment tanks," *Int J Syst Evol Microbiol*, vol. 62, no. 9, 2012, doi: 10.1099/ijs.0.032342-0.
- [60] J. G. Lim and D. H. Park, "Degradation of polyvinyl alcohol by Brevibacillus laterosporus: Metabolic pathway of polyvinyl alcohol to acetate," *J Microbiol Biotechnol*, vol. 11, no. 6, 2001.
- [61] G. Li, Z. Liang, T. An, Z. Zhang, and X. Chen, "Efficient bio-deodorization of thioanisole by a novel bacterium Brevibacillus borstelensis GIGAN1 immobilized onto different parking materials in twin biotrickling filter," *Bioresour Technol*, vol. 182, 2015, doi: 10.1016/j.biortech.2015.01.120.
- [62] M. Abu Talha, M. Goswami, B. S. Giri, A. Sharma, B. N. Rai, and R. S. Singh, "Bioremediation of Congo red dye in immobilized batch and continuous packed bed bioreactor by Brevibacillus parabrevis using coconut shell bio-char," *Bioresour Technol*, vol. 252, 2018, doi: 10.1016/j.biortech.2017.12.081.
- [63] R. K. Mohapatra, S. Pandey, H. Thatoi, and C. R. Panda, "Reduction of chromium(VI) by Marine bacterium brevibacillus laterosporus under varying saline and pH conditions," *Environ Eng Sci*, vol. 34, no. 9, 2017, doi: 10.1089/ees.2016.0627.
- [64] B. ren Yang, Z. qiu Sun, L. ping Wang, Z. xia Li, and C. Ding, "Kinetic analysis and degradation pathway for m-dichlorobenzene removal by Brevibacillus agri DH-1 and its performance in a biotrickling filter," *Bioresour Technol*, vol. 231, 2017, doi: 10.1016/j.biortech.2017.01.038.
- [65] H. N. Nassar, N. S. El-Gendy, M. A. Abo-State, Y. M. Moustafa, H. M. Mahdy, and S. A. El-Temtamy, "Desulfurization of dibenzothiophene by a novel strain Brevibacillus invocatus C19 isolated from Egyptian coke," *Biosci Biotechnol Res Asia*, vol. 10, no. 1, 2013, doi: 10.13005/bbra/1090.
- [66] A. Chebbi *et al.*, "Biodegradation of malodorous thiols by a Brevibacillus sp. strain isolated from a Tunisian phosphate factory," *FEMS Microbiol Lett*, vol. 362, no. 14, 2015, doi: 10.1093/femsle/fnv097.
- [67] S. S. Gomare, G. K. Parshetti, and S. P. Govindwar, "Biodegradation of Malachite Green by Brevibacillus laterosporus MTCC 2298," Water Environment Research, vol. 81, no. 11, 2009, doi: 10.2175/106143009x407357.
- [68] X. Y. Er, T. W. Seow, C. K. Lim, Z. Ibrahim, and S. H. Mat Sarip, "Biological treatment of closed landfill leachate treatment by using Brevibacillus panacihumi strain ZB1," in IOP Conference Series: Earth and Environmental Science, 2018. doi: 10.1088/1755-1315/140/1/012012.
- [69] K. Wei, H. Yin, H. Peng, G. Lu, and Z. Dang, "Bioremediation of triphenyl phosphate by Brevibacillus brevis: Degradation characteristics and role of cytochrome P450 monooxygenase," *Science* of the Total Environment, vol. 627, 2018, doi: 10.1016/j.scitotenv.2018.02.028.
- [70] R. Hooda, N. K. Bhardwaj, and P. Singh, "Brevibacillus parabrevis MTCC 12105: a potential bacterium for pulp and paper effluent degradation," *World J Microbiol Biotechnol*, vol. 34, no. 2, 2018, doi: 10.1007/s11274-018-2414-y.

[71]	N. Jebril, R. Boden, and C. Braungardt, "The isolation and identification of cadmium-resistant
	Brevibacillus agri C15," in Journal of Physics: Conference Series, 2021. doi: 10.1088/1742-
	6596/1879/2/022015.
[72]	N. Jebril, R. Boden, and C. Braungardt, "Cadmium removal with mutant Brevibacillus Agri C15
	CdRentrapped in calcium alginate gel: Multi-constituent ionic exchange," in AIP Conference Pro-
	<i>ceedings,</i> 2023. doi: 10.1063/5.0150816.
[73]	N. Jebril, R. Boden, and C. Braungardt, "Determination of minimal inhibitory concentration of
	cadmium for Brevibacillus agri C15 and Brevibacillus agri C15 Cdr," in AIP Conference Proceedings,
	2023. doi: 10.1063/5.0150818.
[74]	N. Jebril, R. Boden, and C. Braungardt, "Cadmium Removal with Mutant Brevibacillus Agri C15
	CdrEntrapped in Calcium Alginate Gel: A New Process," in IOP Conference Series: Earth and Envi-
	ronmental Science, 2021. doi: 10.1088/1755-1315/761/1/012028.
[75]	N. Jebril, R. Boden, and C. Braungardt, "Use of ultraviolet-light mutagenesis to generate a mutant

[75] N. Jebril, R. Boden, and C. Braungardt, "Use of ultraviolet-light mutagenesis to generate a mutant with elevated cadmium resistance, B. agri C15 CdR," in *Journal of Physics: Conference Series*, 2021. doi: 10.1088/1742-6596/1879/2/022043.